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Brief Communication

Update on Microbiological and Epidemiological characteristics of *Neisseria meningitidis*

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Microbiology of *Neisseria meningitidis*

Neisseria meningitidis (Meningococcus) is a gram-negative diplococcus, bean shaped, non-motile, non-spore forming, and oxidase positive bacteria. Like other gram-negative bacteria, meningococcus is surrounded by outer membrane lipids, outer membrane proteins (OMPs), and endotoxic lipopolysaccharides (LPS). Moreover, pathogenic meningococci are enveloped by a polysaccharide capsule attached to the outer membrane. Meningococci can grow on most non-selective routine laboratory media such as blood and chocolate agar in microaerophilic condition (5% O₂ and 10% CO₂). This microaerophilic condition can be achieved by using either candle jar or gas generating kits. For rapid diagnosis and in cases in which cultures are negative because of previous antimicrobial therapy, meningococcal capsular polysaccharide antigen can be detected in CSF, blood, and urine by latex agglutination procedures.

Meningococci are exclusively human pathogens and reveal more genetic diversity than most other pathogenic human bacteria. This is partly explained by horizontal intraspecies recombination and incorporation from closely related *Neisseria* species (1). Traditionally, strains were typed by using group specific antisera that recognize surface exposed epitopes on the capsule or the outer membrane. By this method, 13 serogroups (based on capsular polysaccharide), 20 serotypes (based on outer membrane proteins) have been identified (2). Further additional typing is also possible by using the antigenic properties of immunoglobulin A1 (IgA1) proteases and pili (3).

Serotyping is of great importance for the development of vaccination strategies. However, although phenotyping characterization may reveal close genetic relatedness, serotyping is not suitable for modern epidemiological purposes (1). Typing schemes, based on variation of a few genes which are probably under selection pressure will not identify the overall relatedness of the chromosomal genome of *N. meningitidis* (1). By using genetic approaches, in particular multilocus enzyme electrophoresis (MEE), which identifies naturally occurring allelic variations in multiple chromosomal house-keeping genes, a better insight into the epidemiology and clonal expansion of disease causing *N. meningitidis* can be gained (4). Other techniques which can be used for phenotyping characterization of *N. meningitidis* are DNA finger printing and PCR (5,6).

Epidemiology

The only natural reservoir of *Neisseria meningitidis* is the human nasopharyngeal mucosa and approximately 10% of the human populations harbor meningococci in the nose. Meningococci are transferred from one person to another by close contact or via respiratory droplets for a distance up to one meter (7). However, invasive disease caused by meningococci is relatively rare, as it occurs only

the following conditions have to be fulfilled (8). These conditions are (i) exposure to a virulent strain, (ii) colonization of the naso-oropharyngeal mucosa by virulent strain, (iii) penetration of the bacterium through the mucosa, and (iv) survival and multiplication of the meningococcus in the bloodstream. These properties are influenced by bacterial properties, climatological and social conditions, preceding or concomitant viral infections, the immune status, and age of the patient.

Meningococcal disease occurs world-wide as endemic infection (1, 8-10). Serogroups B and C cause the majority of infections in developed countries, while serogroups A and, to a lesser extent, C dominate in developing countries (1,8,-11). The incidence of meningococcal disease during the last 30 years varied from 1-3/100,000 population in most developed countries to 10-25/100,000 populations in some developing countries. These different attack rates reflect the different pathogenic properties of *N. meningitidis* strains and different socioeconomic, environmental, and climatological conditions.

Sub-Saharan African countries have a special epidemiological pattern. This region, designated as "meningitis belt", which extends from Mali across the semiarid Sahel zone south of the Sahara, was first described by Lapeyssonnie in 1963. The countries included in this belt are Burkina Faso, Ghana, Togo, Benin, Niger, Nigeria, Chad, Cameroon, Central African Republic, and the Sudan (12). Later, Ethiopia, Mali, Guinea, Senegal, and the Gambia were added to form what is presently denoted as "the expanded meningitis belt" (9,13). In this region, meningococcal disease caused by serogroup A occurs in periodic recurrent waves. The disease attack rate rises at the end of the dry season and declines rapidly after the beginning of the rainy season (8-10, 12). During epidemic peaks, the incidence rate of the diseases may approach 1,000/100,000 inhabitants (9). Initially, a cyclic pattern with epidemics every 8-12 years was reported, but this pattern has

not been confirmed in later studies for most of the countries (9, 12, 15).

Since the end of the 1960s, widespread epidemics due to genetically closely related strains of *N. meningitidis* belonging to clonal complexes have occurred (1).

The biggest epidemics, which originated in Northern China and spread to the south and later globally, were caused by clones of serogroup A (1, 10). These subgroups spread to the Indian subcontinent in 1983 to 1987. In 1987, this clone reached the Middle East and caused a massive epidemic among pilgrims during the Hajj in Mecca (1, 8,9,10). From here the organism was transported with the Hajjis (14), causing epidemics in 1988 in the Sudan and Chad and, in the following years, in Ethiopia, Kenya, and Uganda (15). In the 1990s, the epidemic moved to countries south of the traditional meningitis belt, reaching Nigeria and South Africa in 1996 (16). During this period, more than 150,000 cases and, at least, 16,000 deaths were reported in Africa (1, 10, 17). Interestingly, transfer of strains from the same clonal complex by Hajjis to the United States and Europe did not elicit epidemics in these parts of the world (14, 18).

In Ethiopia, meningococcal meningitis was reported for the first time in 1901, followed by outbreaks reported in 1935, in the 1940s and 1950s, in 1964, 1977, and in 1981 - 1983 and 1988-89 (19). The earlier epidemics were thought to have spread from West Africa to Ethiopia, while the 1988-89 epidemic spread with pilgrims returning from Mecca (15). Bacteriologic studies during the 1981-83

epidemic period showed that serogroups A and C were dominating (20) while, the during 1988-89 epidemic, serogroup A (with one isolate of B) was the most prevalent serogroup (11,21-22). Serogroup A was the cause of the reported recent epidemic of meningitis in Ethiopia (personal communication).

Conclusion

In conclusion, microbiological and epidemiological studies by modern molecular methods have disclosed a complex picture of a few pathogenic meningococcal clones spreading world-wide. However, the mechanism by which potential pathogenic strains cause large-scale epidemics only in some regions of the world is largely unknown (14). It appears that the occurrence of invasive meningococcal disease is not determined solely by the introduction of a new virulent bacterial strain but also by other factors that enhance transmission and by the susceptibility of the population (23). Having a knowledge of an update information related to the microbiological and epidemiological characteristics of meningococci provide basic information for microbiologists, epidemiologists and clinicians to design appropriate diagnostic, preventive, and curative measures, respectively, to the populations which are at risk, particularly the young children which are primarily the victims of this disease.

References

1. Caugant DA. Population genetics and molecular biology of *Neisseria meningitidis*. *APMIS*. 1998; 106:505-525.
2. Frasch C, Zollinger WD, Poolman JT. Serotype antigens of *Neisseria meningitidis* and a proposed scheme for designation of serotypes. *Rev Infect Dis* 1985; 7:504-510.
3. Stephens DS, Whitney AM, Rothbard J, Schoolnik GK. Pili of *Neisseria meningitidis*. Analysis of structure and investigation of structural and antigenic relationships to gonococcal pili. *J Exp Med*. 1985; 161:1539-1553.
4. Selander RK, Caugant DA, Ochman H, Musser JM, Gilmour MN, Whittam TS. Methods of multilocus electrophoresis for bacterial population genetics and systematics. *Appl Environ Microbiol*. 1986; 51:873-884.
5. Woods JP, Kersulyte D, Tolan RW, Berg CM, Berg CE. Use of arbitrarily primed polymerase chain reaction analysis to type disease and carrier strains of *Neisseria meningitidis* isolated during a university outbreak. *J Infect Dis*. 1994; 169:1384-1389.
6. Bjorvatan B, Hassan-King M, Greenwood R, Haimanot RT, Fekade D, Sperber G. DNA fingerprinting in the epidemiology of African Serogroup A *Neisseria meningitidis*. *Scand J Infect Dis*. 1992;24:323-332.
7. Nelson JD. Jails, microbes, and the three foot-barrier. *N Engl. J Med* 1996; 335:885-886.
8. Schwartz B, Moore PS, Broome CV. Global epidemiology of meningococcal disease. *Cin Microbiol Rev*. 1989;2 (Suppl.): S118-S124.
9. Riedo FX, Plikaytis BD, Broome CV. Epidemiology and prevention of meningococcal disease. *Pediatr Infect Dis J* 1995; 14:643-657.
10. Achtman M. Global epidemiology of meningococcal disease. In K. Cartwright (ed), *Meningococcal disease*. P. 159-175. John Wiley & Sons, Ltd., Chichester, United Kingdom, 1995.
11. Tekle Haimanot R, Caugant DA, Fekadu D, et al. Characteristics of serogroup A *Neisseria meningitidis* responsible for an epidemic in Ethiopia, 1988-89. *Scand J Infect Dis* 1990;22:171-174.
12. Lapeyssonnie L. La meningite cerebro-spinale en Afrique. *Bull. W.h.O.* 1963;28 (Suppl. 1): 3-14.
13. Greenwood BM, The epidemiology of acute bacterial meningitis in tropical Africa, P 61-69, *Bacterial meningitis*. Academic Press, Ltd., London, United Kingdom, 1987.

14. Moore PS, Hierholzer, Dewitt W, Gouan K, Djore D, Lippeved T, Plikaytis B, Broome CV. Intercontinental spread of an epidemic group A *Neisseria Meningitidis* strain. *Lancet* ii. 260-263.
15. Moore PS. Meningococcal meningitis in sub-Saharan Africa: a model for the epidemic process. *Clin Infect Dis*. 1992;14:515-525.
16. McGee L, Koornhof HJ, Caugant DA. Epidemic spread of subgroup III of *Neisseria meningitidis* serogroup A to South Africa in 1996. *Clin Infect Dis* 1998;27:1214-1220.
17. Guibourdenche ME, Hoiby EA, Riou jY, Varaine F, Joguet C, Caugant DA. Epidemics of serogroup A *Neisseria meningitidis* of subgroup III in Africa, 1989-94. *Epidemiol Infect* 1996;116:115-120
18. Moore PS, Harrison LH, Telzak E, Ajello GW, Broome CV. Group A meningococcal carriage in travelers returning from Saudi Arabia. *JAMA*. 1988;260:2686-2689.
19. Tadele T. Meningococcal meningitis. In: Helmut Kloos and Zein Ahmed Zein (eds.), the ecology of health and disease in Ethiopia p285-293, westview press 1993.
20. Estifanos T, Habte-Gabr E, Gebre-Yohannes A. Meningococcal meningitis. In Zein Ahmed Zein and H. Kloos (ed.), *The Ecology of Health and Disease in Ethiopia*. P273-280, Addis Ababa: Ministry of Health, 1988.
21. Hariga F. Cerebrospinal meningitis epidemic and surveillance system in the Sudan, Ethiopia and Chad. WHO, Pan African Centre for Emergency preparedness and Response, Addis Ababa, 1990.
22. Habte-Gabr E, Estifanos T, Mamao M. Meningococcal meningitis in Ethiopia. *Ethiop J Hlth Dev* 1984;1:47-63.
23. Stephens DS. Uncloaking the meningococcus: dynamics of carriage and disease. *Lancet*. 1999;353:941-942.