

The Ethiopian Journal of Health Development

Original article

Isolation and characterization of *Cryptococcus neoformans* from environmental sources in Ethiopia

Yimtubezenash W/Amanuel¹, Leykun Jemaneh¹, Dawit Abate²

¹Department of Medical Microbiology and Parasitology, Faculty of Medicine, Addis Ababa University P.O. Box 9086, Addis Ababa, Ethiopia; ²Department of Biology, Faculty of Science, Addis Ababa University, Addis Ababa, Ethiopia

Ethiop. J. Health Dev. 2001;15:45-50

Abstract

Background: *Cryptococcus neoformans*, the most important cause of fungal meningitis in immunocompromised patients worldwide has two varieties which differ in their geographical distribution and natural habitat.

Objective: To isolate and characterize *C. neoformans* from environmental sources in and around Addis Ababa.

Method: A total of 592 specimens collected from droppings of pigeons and chickens, and from material associated with eucalyptus trees were examined for the presence of *C. neoformans*. This study was carried out during September 1998 to July 1999.

Results: Out of the 592 specimens eleven *C. neoformans* were isolated from the pigeon droppings and material associated with eucalyptus trees. No *C. neoformans* was discovered from chicken droppings. All the isolates were identified as *C. neoformans* var. *neoformans*. *C. neoformans* var. *gattii* has not been isolated from any of the samples examined.

Conclusion: The fact that cultural detection of *C. neoformans* var. *gattii* was impossible in this study does not, however, prove the absence of this variety in these environmental sources in Ethiopia. Future studies that will include a wider geographical area in Ethiopia, and also investigate the possible association of exposure to environmental sources and risk of acquiring disease are recommended. [Ethiop. J. Health Dev. 2001;15(1):45-49]

Introduction

Cryptococcus neoformans (teleomorph, *Filobasidiella neoformans*) is an encapsulated round or oval yeast. It is subdivided into two varieties and four major serotypes, based on capsular epitopes: *C. neoformans* var. *neoformans* (serotypes A and D) and *C. neoformans* var. *gattii* (serotypes B and C) (1). A distinctive difference in the geographic distribution of the two varieties has been reported (2). *C. neoformans* var. *neoformans* occurs worldwide while var. *gattii* is restricted to tropical and subtropical regions (3,4).

In nature the two varieties reside in separate environmental niches, which have been identified as potential sources of pathogenic strains (5). *C. neoformans* var. *neoformans* has been isolated from a variety of avian species in addition to pigeons, including chickens, parrots, sparrows and sites contaminated by pigeon excrement and at a lesser concentration in soil (6,7). *C. neoformans* var. *gattii* has been isolated from *Eucalyptus camaldulensis* (3,4), and from *Eucalyptus tereticornis* (8,9). *Eucalyptus* trees particularly *E. camaldulensis* and *E. globulus* originated from Australia and are extensively grown in many countries (3), including Ethiopia, particularly in and around towns and cities of the highland regions. In Ethiopia *E. globulus* is one of the earliest introduced species, and *E. tereticornis* and *E. camaldulensis* are commonly and widely planted species (10).

The disease caused by variety *neoformans* occurs throughout the world, while cryptococcosis caused by variety *gattii* is found mainly in tropical and subtropical areas (11). *Cryptococcus* has emerged as the fourth most common lethal infection among AIDS patients (12,13). In these patients meningoencephalitis is the most common presentation. The first case of cryptococcal meningitis in Ethiopia was described in 1992 (14). In another related study among 305 seropositive Ethiopian patients 28 developed cryptococcal meningitis (15).

The clinical isolates from AIDS patients have been identified predominantly as *C. neoformans* var. *neoformans* (16). Similar results have been obtained in Ethiopian HIV-positive patients, where all the isolates from patients were identified as serotype A (17). Isolation of strains of the variety *gattii* from patients with AIDS is rarely reported (18,19,20,21). It has been observed that the clinical course was prolonged, and morbidity, mortality, and severe neurological sequelae were more frequent among patients infected with *C. neoformans* var. *gattii* (22).

In Ethiopia, it is common to have chicken and pigeon droppings, and additionally eucalyptus trees around the living areas, and the possible exposure to infection is wide.

In this paper we report the first environmental isolation of *Cryptococcus neoformans* from different natural sources in and around Addis Ababa.

Methods

A total of 592 samples, 200(flower), 55(wet leaf), 77 (dry leaf) from Eucalyptus trees; 100 (dry), 92 (fresh) droppings of pigeon; and 20 (dry), 48 (fresh) chicken droppings were collected between September 1998 to July 1999. All the samples were from different sites in Addis Ababa and its surroundings.

The samples from eucalyptus trees were transported to the laboratory in clean dry paper envelopes. The samples were mixed with 25 ml sterile distilled water in 250 ml Erlenmeyer flasks, which were then shaken vigorously for one hour, and the contents allowed to settle for 15 min (23). From the supernatant 0.1 ml aliquot was aseptically aspirated and plated on to plates containing *Guizota abyssinica* creatinine agar (Bird seed agar) (24) This medium is differential for this yeast as it gives pigmented colonies. The medium was supplemented with chloramphenicol to suppress bacterial growth. The cultures were incubated at 37°C and were observed daily for the development of brown yeast-like colonies.

Pigeon droppings were obtained from underneath buildings where pigeons are common. Chicken droppings (fresh and dry) were collected in sterile tubes from market places in Addis Ababa. The samples were then suspended in sterile water and shaken. The suspension is allowed to settle and aliquotes were plated on bird seed agar (BSA) and incubated at 37°C.

The plates were then observed for appearance of brown colonies. All suspected colonies were subcultured on Sabouraud glucose agar and india ink preparations were made. The isolates were further identified on the basis of a positive India ink test, urease production, growth at 37°C, and by carbohydrate assimilation and fermentation tests by API 20C (bioMerieux, Hazelwood, Mo.) testing (25).

The positive isolates which were confirmed to be *C. neoformans*, were further tested on canavanineglycine-bromthymol blue (CGB) agar for the differentiation of *C. neoformans* var. *neoformans* (serotypes A/D) and *Cryptococcus neoformans* var. *gattii* (serotypes B/C) (1).

Reference strains *C. neoformans* CBS 5757 and *C. neoformans* CBS 5756 obtained from Karolinska Institute and Hospital, Department of Clinical Microbiology, Sweden, were used as control throughout the study.

Results

Of the total 592 samples, brown-like colonies were detected on Bird seed agar on eleven cultures. These were found to be India Ink positive, urease positive and grew at 37°C and gave characteristic carbohydrate assimilation profile, showing the reliability of the bird seed agar for isolation of *C. neoformans*. Samples of pigeon droppings yielded 5% (9/192) positivity, and samples related to eucalyptus trees yielded 0.6% (2/332) positivity (Table 1).

All the isolates identified as *C. neoformans* gave negative test on canavanine-glycine-bromthymol blue (CGB) agar, indicating that the isolates are *Cryptococcus neoformans* var. *neoformans* (serotype A/D). From the total sample, eleven (2%) *Cryptococcus neoformans* var. *neoformans* were obtained. *C. neoformans* var. *gattii* has not been isolated from any of the samples investigated.

Discussion

The isolation of *C. neoformans* in nature took place for the first time from peach juice in 1895 and then from milk in 1901 (26). Then Emmons isolated the organism from soil and then from pigeon excreta in 1950s (27). Since then the two varieties have been isolated from different environmental sources.

This is the first isolation of *Cryptococcus neoformans* from environmental sources in Ethiopia. The percentage of positivity is well within the range reported in the literature: 0 to 24.7% (2,18,28). Nine out of the total isolates were isolated from pigeon droppings. Even though birds do not get infected because of their relative high body temperature, avian droppings are considered an optimal natural substratum for *C. neoformans* due to their high content of low molecular nitrogenous compounds such as creatinine (2,25).

We were unable to isolate the variety *gattii* in this study. This is in accordance with the preliminary work done from HIV positive patients in Ethiopia that no *C. neoformans* var. *gattii* has been isolated (17). The presence of var. *gattii* was expected, since the natural habitat of this variety has been identified as eucalyptus trees which are found abundantly in Addis Ababa and its surroundings. Only five isolates of *C. neoformans* var. *gattii* were obtained from 696 specimens collected from different types of eucalyptus trees in India (6). Therefore, further more extensive studies are needed to confirm the presence or absence of variety *gattii* in Ethiopia.

Since *C. neoformans* is ubiquitous in the environment, exposure to dry inhalable yeasts from this reservoirs is expected. The small dry basidiospores would make effective infectious particles, and they have been suggested to be a source of infection (29). Even though there is a lack of direct evidence that suggests that exposure to these environmental sources is associated with an increased risk of acquiring disease there are some evidences that show that these environmental sources might be potential sources of pathogenic strains. The need of exposure to *E. camaludulensis* trees in order to acquire an infection by the variant *gattii* has been suggested as a plausible explanation for the high incidence of infections caused by this fungus in Australian aborigines and low worldwide incidence of

this serotype in AIDS patients (3). Recently it has been shown that pigeon droppings contained a genetically heterogenous population of *C. neoformans* serotypes A and D in which some isolates are similar to infection-causing organisms demonstrating that pigeon droppings can be identified as potential source of pathogenic strains of *C. neoformans* (5). In previous studies genetically related isolates were found among clinical and environmental strains, suggesting that pathogenic strains of *C. neoformans* can be found in the environments of patients at risk for cryptococcosis (30,31). Since *C. neoformans* is an important pathogen in immunocompromised patients the elucidation of the chain of infection is very important.

Future studies are needed to investigate the geographic distribution of this yeast in Ethiopia, the prevalence of disease caused by this fungus, and possible direct association of disease with exposure to these natural habitats of *C. neoformans*.

Acknowledgments

This work was supported by the grant obtained from Akilu Lemma Foundation. We thank the technical assistance of Ato Biruk Jemaneh.

Tables

Table 1: *C. neoformans* isolated from different environmental sources in and around Addis Ababa

	Samples studied		
	Number	Positive	
		No.	%
Pigeon droppings	192	9	5
Chickens droppings	68	0	-
Eucalyptus trees	332	2	0.6
Total	592	11*	2

*All *C. neoformans* var *neoformans*

Figures

References

1. Kwon-Chung KJ, Polachek I, and Bennet JE. Improved diagnostic medium for separation of *C. neoformans* var. *neoformans* (ser. &D) and *C. neoformans* var. *gatii* (ser. &C). J Clin Microbiol. 1982;15:535-537.
2. Lazera MS, Wanke B, Nishikawa MM. Isolation of both varieties of *Cryptococcus neoformans* from saprophytic sources in the city of Rio de Janeiro, Brazil. J Med Vet Mycol. 1993;31:449-454.

3. Ellis DH, and Pfeiffer TJ. Natural habitat of *Cryptococcus neoformans* var. *gattii*. *J Clin Microbiol*. 1990;28:(7) 1642-1644.
4. Pfeiffer T, and Ellis D. Environmental isolation of *C. neoformans* var. *gattii* from California. *J Infect Dis*. 1991;163:939-930.
5. Garcia-Hermos D, Mathoulin-Pelissier S, Couprie B, Ronin O, Dupont B, Dromer F. DNA typing suggests pigeon droppings as a source of pathogenic *Cryptococcus neoformans* serotype D. *J Clin Microbiol* 1997;35:2683-2685.
6. Littman M L, Borok R. Relation of the pigeon to cryptococcosis: natural carrier state, heat resistance and survival of *Cryptococcus neoformans*. *Mycopathologia*.1968;35:329-45.
7. Levitz SM. The ecology of *Cryptococcus neoformans* and the epidemiology of cryptococcosis. *Rev Infect Dis*.1991;13:1163-1169.
8. Currie B, Vigus T, Leach G, and Dweyer B. *C. neoformans* var. *gattii*. *Lancet*. 1990;336:1442.
9. Padhye AA, Chakrabarti A, Chander J, and Kaufman L. *Cryptococcus neoformans* var. *gattii* in India. *J Med Vet Mycology*. 1993;31:165-168.
10. Flora of Ethiopia and Eritrea. Canllaceae to Euphorbiaceae. eds Sue Edwards, Mesfin Tadesse and Inga Hedberg. Addis Ababa & Uppsala. 1995: vol 2 Part2.
11. Kwon-Chung KJ, and Bennet JE. Epidemiologic differences between the two varieties of *C. neoformans* . *Am J Epidemiol*. 1984;120:123-140.
12. Dissmukes WE. Cryptococcal meningitis in patients with AIDS. *J Infect Dis*. 1988;157:624-628.
13. Mitchell TG, Perferct JR. Cryptococcosis in the Era of AIDS-100 years after the discovery of *Cryptococcus neoformans*. *Clin Microbiol Rev* 1995;8:515-548.
14. Amha G. Cryptococcal meningitis in a young Ethiopian woman with AIDS. *Ethiop Med J* 1992;30:169-73.
15. Teshale S, Alemu S, Assefa A. Cryptococcal meningitis in patients with AIDS (abstract). *Ethiop Med J* 1996;34:258.
16. Rozenbaum R, Goncalves AJR, Wanke B, et al. *Cryptococcus neoformans* varieties as agents of cryptococcosis in Brazil. *Myco-pathologica* 1992;119:133-6.
17. Yimtubezinash WA, Dawit A. *Cryptococcus neoformans* isolated from HIV-positive patients with meningitis(abstract). *EMJ*, 1996;34:259.
18. Swinne D, Nkurikiyinfura JB, Muyembe TL. Clinical isolates of *Cryptococcus neoformans* from Zaire. *Eur J Clin Microbiol*. 1986;5:50-51.
19. Rinaldi MG, Drutz DJ, Howell A, Sande MA, Wofsy CB, Hadley WK. Serotypes of *Cryptococcus neoformans* in patients with AIDs [letter]. *J Infect Dis* 1986;153:642.
20. Bottone EJ, Salkin IF, Hurd NJ, Wormser GP. Serogroup distribution of *Cryptococcus neoformans* in patients with AIDS [letter]. *J Infect Dis*. 1987;156:242.
21. Clancy MN, Fleischmann J, Howard DH, Kwon-chung KJ, Shimizu RY. Isolation of *Cryptococcus neoformnas gattii* fom a patient with AIDS in Southern California [letter].*J Infect Dis* 1990;161:809.
22. Mitchell DH, Sorrell TC, Allworth AM, et al. Cryptococcal disease of the CNS in immunocompetent hosts: influence of cryptococcal variety on clinical manifestations and outcome. *Clin Infect Dis* 1995;20:611-6.
23. Chakrabarti A, Jatana M, Kumar P, Chatha L, Kaushal A, Padhye A. Isolation of *Cryptococcus neoformans* var.*gattii* from *Eucalyptus camaldulensis* in India. *J Clin Microbiol*. 1997;3:3340-3342.
24. Evans EGV, Richardson MD. 1989. *Medical mycology, a practical approach*. IRL press, Oxford University press, Oxford.
25. Kwon-Chung KJ & Bennett JE. 1992. *Medical Mycology* Lea and Febiger, Philadelphia, Pa.

26. Rippon JW. Medical Mycology. The pathogenic fungi and the pathogenic actinomycetes. 3rd ed. Philadelphia: W. B. Saunders, 1988:582-609.
 27. Emmons CW. Saprophytic sources of *Cryptococcus neoformans* associated with the pigeon (*Columba livia*). Am J Hyg. 1955;62:227-32.
 28. Ruiz A, Velez D, Fromtling RA. Isolation of saprophytic *Cryptococcus neoformans* from Puerto Rico: Distribution and variety. Mycopathologia 1989;106:167-170.
 29. Ellis DH, Pfeiffer TJ. The ecology of *Cryptococcus neoformans*. Eur J Epidemiol. 1992;8:321-325.
 30. Currie Bp, freundlich LF, Casadevall. Restriction fragment length polymorphism analysis of *C. neoformans* isolates from environmental (pigeon excreta) and clinical sources in New York City. J Clin Microbiol 1994;32:1188-1192.
 31. Yamamoto Y, Kohno S, Koga H et al. Random amplified polymorphic DNA analysis of clinically & environmentally isolated *C. neoformans* in Nagasaki. J Clin Microbiol 1995;33:3328-3332.
- 